

## **EXHIBIT C**

TITLE <sup>32</sup>P-Kinasing of ChtA176Project No. \_\_\_\_\_  
Book No. 391

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Purpose - To label synthetic probe ChtA176 with <sup>32</sup>P by kinasing for Chlamydia assay development.

Reagents -

Synthetic probe ChtA176 mm 320-39 1.28 OD/ml 64 µg/ml  
 10X Kinase buffer, from 2B + mm, aliquot obtained (391:18)  
 T4 Kinase, BRL, 10 µl/pel lot 62111  
 Y-<sup>32</sup>P-ATP 3000 Ci/mmol # G551, acd 10 µCi/pel  
 Tutton, acid base treated, from Frank H.  
 5 m NaCl DES NB 157:48  
 1% SDS diluted from 20% (DK)  
 1 M Tris, pH 8.2 (DK)  
 100 mM EG 1/N 500:38  
 4 M LiCl from DK  
 Proteinase K 20 mg/ml 333:11  
 Glycogen M-1 ~40 mg/ml in 10% EtOH (DK)  
 Phenol from DK ( )  
 Chloroform  
 10% TCA  
 OSA, 10 mg/ml from BRL Lot # 40416

## Procedure

- Follow same procedure used (391:18-20)
- Kinase reaction included 1 µl of ChtA176 (64 ng)

## Results

Column areas in 0.3 m NaCl and H<sub>2</sub>O :

$$\Sigma (9.6 \times 10^4) \left( \frac{3300}{5} \right) = 6.3 \times 10^7 \text{ cpm}$$

Straight count -  $(6.2 \times 10^4) \left( \frac{200}{1} \right) \left( \frac{50}{10} \right) = 6.2 \times 10^7 \text{ cpm}$  → why so low?

Input dpm =  $2.2 \times 10^8 \text{ dpm}$

EtOH super -  $(1.1 \times 10^3) \left( \frac{2400}{5} \right) = 5.3 \times 10^5 \text{ cpm}$

Witnessed &amp; Understood by me,

John Kop

Date

Invented by

Margie Harper

Recorded by

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Date

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$$\text{TCI applicable cpm} = (1.2 \times 10^4) \left( \frac{200}{1} \right) \left( \frac{50}{10} \right) = 1.2 \times 10^7 \text{ cpm in } 64 \text{ ng}$$

specific act. =  $1.9 \times 10^8 \text{ cpm}/\mu\text{g}$

$$\text{Final cpm} \quad (1.8 \times 10^9) \left( \frac{100}{2} \right) = 9 \times 10^6 \text{ cpm} \quad 75\% \text{ recovery}$$

$9 \times 10^4 \text{ cpm}/\mu\text{l}$

Mixed total sample to 300  $\mu\text{l}$  0.02% SDS  $\rightarrow 3 \times 10^4 \text{ cpm}/\mu\text{l}$  final  
store at 4°C

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